

Influence of Environmental Factors on the Mosquito Vector Habitat Distribution in Urban Areas of Faisalabad, Punjab

Sania Shamas^{1,2,#}, Maryam Riasat^{2,#}, Rida Younas², Naureen Rana², Nawaz Haider Bashir¹, Muhammad Naeem¹, and Huanhuan Chen^{1,*}

¹ College of Biological Resource and Food Engineering, Qujing Normal University, Qujing 655011, China.

² Department of Zoology, Faculty of Engineering and Applied Sciences, Riphah International University, Faisalabad Campus, Faisalabad, 38000, Pakistan.

*Correspondence: chhuanhuan@163.com

Article Info

Academic Editor: Saba Malik

Received: 25, May, 2025

Accepted: 31, May, 2025

Published: 1 July, 2025

Citation: Shamas, S., Riasat, M., Younas, R., Rana, N., Bashir, N. H., Naeem, M., & Chen, H. (2023). Influence of environmental factors on the mosquito vector habitat distribution in urban areas of Faisalabad, Punjab. *Pakistan Journal of Zoological Sciences*, 1(1), 1-14.

Copyright: © 2025 by the authors. This article is submitted for possible open access publication under the terms and conditions of the [Creative Commons Attribution \(CC BY\) license](https://creativecommons.org/licenses/by/4.0/).

© 2025 IJSMART Publishing Company. All rights reserved.

Abstract The viruses that cause malaria, Dengue hemorrhagic fever, and Rift Valley fever are primarily spread by mosquitoes. Globally, mosquito-borne diseases pose a serious threat to public health, particularly in crowded cities. The goal of the current study was to update knowledge about the mosquito (Diptera: Culicidae) fauna of Punjab region, Pakistan's District Faisalabad, and forecast the distribution of the larvae of the most important mosquito vectors in this area. Environmental factors such as water sources, land use, temperature, and humidity were recorded at each collection location. A perspective for the geographic distribution of dengue vectors in the metropolitan areas of Faisalabad was created using GIS-based spatial analytic tools after the data was gathered. The number of mosquito larvae was assessed in connection with the physiochemical characteristics (pH & TDS) of breeding grounds. Mosquito larvae were collected from January 2024 until December 2024. *Aedes aegypti* was the most important vector discovered in Faisalabad. To predict the species distribution of *Aedes aegypti* in the district of Faisalabad, 19 bioclimatic variables were combined with the data from the collection. More than 100 locations yielded more than 1800 mosquito larvae. The northeastern region of the Faisalabad district was identified as having the best suited lands. The region that was least conducive to the presence of *Aedes aegypti* was the southwest. Precipitation factor (bio8) contributed the most in the presence of this vector, accounting for over 40%, followed by bio19, which contributed 20%.

Keywords: Mosquito vectors, spatial distribution, GIS, dengue, *Aedes aguptii*, urban areas, Faisalabad, Punjab, Pakistan.

Introduction

Mosquito-borne diseases are a growing concern in many urban areas, especially in developing countries like Pakistan. In cities such as Faisalabad, rapid urban growth, poor drainage, stagnant water, and changing weather conditions create ideal breeding grounds for mosquitoes. Factors like temperature, humidity, rainfall, vegetation, and land use play a key role in shaping where mosquitoes can live and thrive. This study aims to explore how these environmental elements influence mosquito habitats across different parts of Faisalabad. Using mapping tools and

ecological models, the research will help identify areas at greater risk, ultimately supporting more focused and effective mosquito control and public health strategies.

Due to the change in the climate, urbanization and human activities, Pakistan has to deal with high risk of mosquito-borne illnesses in last few years. In disease transmission, researches from region such as Peshawar and Punjab have highlighted the role of *Aedes* mosquitoes, particularly *Ae. aegypti* and *Ae. albopictus*. Research indicates that these species differ in bacterial diversity and *Wolbachia* infection, with *Ae. albopictus* hosting more diverse strains. Environmental surveys have recognized parks,

#Both authors contributed equally.

forests, and scrapyards as key breeding grounds, with mosquito diversity peaking during the rainy season. Advanced tools like GIS and AHP have also been used to map high-risk areas and assess environmental factors. These insights support integrated, targeted mosquito control strategies by combining ecological, microbial, and spatial data.

Ammar et al. (2025), discovered that due to climatic changes in Pakistan, deforestation, urbanization and poor sanitation causes the arboviral diseases at high level. As there is no proper investigation system in Pakistan, it causes the great hurdle in the way effectively tracking the disease prevalence. High prevalence rate of dengue, West Nile virus, chikungunya, and Japanese encephalitis have caused the major health issues along with other hidden arboviral diseases. So improved investigation system, research and control strategies are required to overcome these challenges.

Nayab et al. (2025), completed the research work by focusing on *Ae. aegypti* and *Ae. albopictus*. Mosquitoes that were collected using Gravitraps, and their species were confirmed through molecular methods, these investigations have pointed out the presence of Wolbachia in *Aedes* mosquitoes, in Peshawar, the northwest region of Pakistan. With *Ae. albopictus* hosting 921 bacterial species, while *Ae. aegypti* had only 239 species, the 16S rRNA sequencing disclosed remarkable differences in bacterial diversity. Both species were found to carry Wolbachia, with *Ae. aegypti* infected with Wolbachia pipientis, and *Ae. albopictus* showing a co-infection of two Wolbachia strains, Wolbachia pipientis and Wolbachia bourtzisii. To control dengue and other mosquito-borne diseases in the area, these researches pointed out the significance for using Wolbachia.

Mehmood et al. (2024), found that, based on behavioral and feeding habit, mosquito species show the great degree of variations in habitat selection. By analyzing microhabitats, species diversity, and habitat web structures through the surveys conducted in the Chakwal district of Punjab, Pakistan, identified the potential mosquito's habitats. A total of 580 mosquito specimens were collected, representing 12 species from five genera. Crop fields had the lowest while parks, forest areas, and scrapyards had the highest mosquito populations. Parks also emerged as the most species-rich habitats, with graveyards showing the least diversity.

Ullah et al. (2023) explored that Mosquito distribution across different agro-ecological zones of Punjab is shaped by seasonal changes, land use, and human activity, all of which influence disease risk. Surveillance was conducted in six zones, collecting samples from stagnant water sources during various seasons. A total of 24 mosquito species were identified, with higher diversity observed during the rainy season. Anopheline mosquitoes were found across both rural and urban areas, while Aedine species were mostly limited to northern irrigated plains, especially around Changa Manga Forest. Culicine mosquitoes were widespread and present

year-round. Some species showed specific habitat preferences, such as *C. crassipes* in rainfed areas and *M. chamberlaini* in both urban and rural environments. Climatic factors significantly influence the presence of *Aedes aegypti* larvae in indoor and outdoor water containers across various districts. Analysis showed a strong link between weather patterns and larval abundance, except in Faisalabad during the summer, where no significant association was found. Containers containing stagnant water placed at specific places also played a significant role in this purpose. These researches provided the remarkable understandings and demonstrations about potential breeding sites of mosquitoes and manipulating the crucial mosquitoes control strategies (Malik et al., 2023).

In 2019, Ali and Ahmed, provided a detailed map that represents the complete sketch of diseases caused by mosquitoes in Kolkata Municipal Corporation, with the help of geographical information system (GIS) and analytical hierarchy technique. To recognize the weightage of chosen items, a pairwise contrasting matrix, a choice-based classifying technique was formulated.

Jemal and Thukair (2018), illustrated that a pairwise comparison matrix was used to assess the weights of ten pertinent environmental components. A geo-database was also created using the weight of each causal element to facilitate overlay analysis. To confirm the significance-measurement and decision-making processes, the consistency ratio was created. Because it is less than 0.1, the compatibility ratio to the select variables, which was determined to be 0.0470, is considered accurate. The study's analysis shows that a number of important factors.

The current study demonstrates the extensive applicability of satellite data and a spatial approach to the zonation of epidemic diseases. Numerous studies across the globe have highlighted the effectiveness of Geographic Information Systems (GIS) and environmental modeling tools in understanding mosquito vector distribution. For instance, research conducted in the Eastern Province of Saudi Arabia used GIS and meteorological data to map mosquito breeding sites, revealing that temperature, humidity, and rainfall play key roles in mosquito abundance. Similar work in Hawaii emphasized the need to include environmental variation—such as rainfall and temperature thresholds—when predicting mosquito habitats, especially in areas with high population density. In Virginia, USA, GIS was used alongside remote sensing and field data to forecast the abundance of mosquito species, showing how environmental factors could guide risk assessments and disease surveillance.

Meanwhile, in Pakistan, although species distribution modeling (SDM) has been used to study mosquitoes, GIS-based studies remain limited. Past outbreaks like the 2017 dengue epidemic in Khyber Pakhtunkhwa highlight the urgent need for spatial modeling. Studies using techniques such as binary logistic drift and binomial kriging have shown promising results in identifying high-risk areas. Traditional mosquito

control methods, such as chemical pesticides, have been challenged by rising resistance in mosquito populations, making ecological modeling even more critical.

Materials and Methods

Description of the study area

Pakistan's terrain is defined by a number of regions, including the Indus Plain, the Northern and North-Western Mountains, the Potwar Plateau, the Balochistan Plateau, and Salt Range, and other ecosystems and climate zones. Faisalabad, Pakistan, is a 3.4 million-person city that occupies 3,358 km² in northeastern Pakistan, situated between the Ravi and Chenab river plains. According to Saleem and Mahmood (2023), the southern half is estimated to be bordered by Sahiwal and Toba Tek Singh, the northern extremity by Chiniot and Hafizabad, Nankana Sahib to the east, and Jhang to the west.

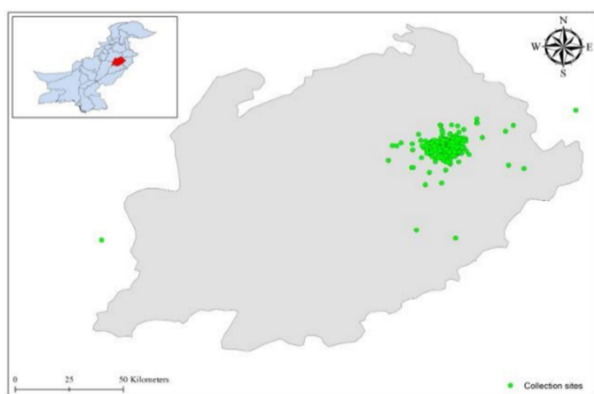


Figure 1. Pakistan map showing collection sites in Faisalabad.3.2.

Data Collection

Field trips and surveillance operations were carried out to collect mosquito larvae. After sampling every breeding location, the most usual and promising habitats were chosen from both lower and higher elevations. From January 2024 to December 2024, the collection of mosquito larvae was conducted continuously. To collect mosquito larvae, a regular 350 ml plastic dipper was utilized. By using a global positioning system unit (GPS), the coordinates of the collection sites were acquired (Garmin, eTrix). Each recovered mosquito larva was identified and submerged in 80% alcohol.

Diverse Breeding Habitats:

Various breeding sites were surveyed for mosquito larvae collection, including streams, wells, water containers, and pools. Here are some images depicting these sites.



Figure 2. Mosquito larvae emerged in the stagnant water present in a tree aperture, Jinnah colony, Faisalabad district.



Figure 3. Rainwater accumulation in a park of Chak Jhumra, Faisalabad district.

Identification of Mosquito

Larvae were gathered from several nesting locations using conventional dipping methods in order to identify mosquito species. They were then raised in a controlled laboratory environment until they matured into adult mosquitoes. Morphological keys and taxonomic guides were then used to identify the emerging adults down to the species level (Rueda, 2004; Gillies & Coetzee, 1987). Important morphological traits such wing venation, leg banding patterns, and abdominal markings were used for identification.

Morphological identification is still a valid way to distinguish between mosquito species, especially in field-based research, according to Alahmed et al. (2011). Their study acknowledged that environmental influences can affect morphological changes, but it also emphasized the significance of defined taxonomic keys for precise species confirmation. In

order to ensure correctness, we used published taxonomic keys and expert assistance to crossverify our identification method. For use in MaxEnt, Dengue mosquitos were recognized and data was monitored in CSV (Comma separated value) format.

Physiochemical characteristics

The pH and total dissolved solids of every breeding locations were analyzed through the pH meters along with TDS (HANNA Instruments, USA).

Visualisation and analysis

The result of molecular docking was analysed using different software such as AutoDock 4.2, PyMOL and LigPlot program. PyMOL program was mainly used for visualisation of the structures. The main parameters for molecular docking analysis were the binding energy values of the protein-ligand interaction, hydrophobic interaction and the number of hydrogen bond formation. The binding energy value for each compound was shown as a result of molecular docking and the best interacting compound of both enzymes was selected. Besides, the number of hydrogen bonds formation and the residues that formed the hydrogen bonds with the ligand were identified using AutoDock 4.2 software. The distance of hydrogen bonding between the ligand and the interacting residue was being calculated using PyMOL. Also, AutoDock 4.2 software was used to reveal all the interacting residues of the protein, and each ligand; meanwhile, Ligplot software was used to visualise the hydrophobic interaction of the protein and the ligands.

Modeling procedures

Accurate test statistics and independent evaluating data is required due to the nichebased nature of the distributional models. In order to offer distinct test data, the Faisalabad district was selected for the collecting of mosquito larvae kinds from multiple grid cells. Test statistics were then acquired using Maxent software. To describe the environment of mosquito larvae, 19 bioclimatic layers temperature layers and According to Phillips et al. (2006), from the WorldClim database version 1.4 (www.worldclim.org) eight precipitation levels were taken. 30 arc-seconds (1 km) is spatial resolution of these bioclimatic strata. After being shrunk to the size of the Faisalabad district, in ASCII grid format each of these bioclimatic layers was recorded so that MaxEnt could use it. The model builder tool in ArcGIS version 10 was used to reduce these layers. The geographical distribution of the species was modeled using MaxEnt software version

3.3. The program was set up using the "Auto features" option, ASCII output file format, and logical output structure as recommended by Phillips and Dudík (2008). The Maxent software connected predictor characteristics (environmental and topographical) with the distribution of target potential species.

The value assigned to any pixel is = (The probabilities of that pixel + probabilities of all other pixels having equal or lower probability values) X 100

Any pixel with a value of 100 indicates the most adaptability to depict the presence of the species, while a value of 0 indicates the least compatibility (Phillips et al., 2004; Phillips et al., 2006).

Evaluation Metrics

MaxEnt

Maximum Entropy (MaxEnt) originated in statistical mechanics (Jaynes, 1957). It is a common method for predicting topographical dispersion of mosquitoes in the research area. Only presence data and ecological data are required for MaxEnt to simulate species distribution (Phillips et al., 2006). In a large-scale model discrimination investigation, Maxent performed well (Elith et al., 2006). Similarly, the analysis of four species distribution modeling approaches revealed that, even with a small sample size, maxent could produce extremely valuable and accurate results (Hernandez et al., 2006).

The model developed using this approach also shows how climate change impacts the habitat of the target species (Phillips et al., 2004; Hernandez et al., 2006). Only a little number of specimens can be employed for the species compatibility referred by maximum (Pearson et al., 2007; Ortega and Townsend, 2008; Wisz et al., 2008; Kija et al., 2013). Even with sample numbers as little as five, Maxent has proven to have a high success rate (Pearson et al., 2007).

The Genetic Algorithm for Rule-set Production (GARP) performs worse than maxent for sample sizes smaller than 10 (Pearson et al., 2007). The marginal density along the study area region contrasted to the conditional density at the presence sites of predictors to forecast species-suitable habitats using maxent (Conley et al., 2014). Maxent can generate species distribution models using five occurrence tracks (Hernandez et al., 2006). In any event, for a reliable forecast, at least 30 recurrence data should be present (Wisz et al., 2008). Models produced by Maxent show less omission errors than those produced by GARP (Foley et al., 2010).

Determining the importance of variables in model building

The relative importance and value of each variable in the geographic distribution of species models are analyzed by Maxent using the Jackknife technique. By the Jackknife test, three different model types are produced (Phillips et al., 2006; Phillips and Dudik, 2008):

- Models created with all but one variable considered.
- Models created with one variable and all other variables eliminated at the same time.
- Models that contained all of the variables.

The initial learning gain increases when the model is developed with only one variable, while decreases as the most important variable is eliminated from the model, but. In other words, the least AUC values shown by variables that are eliminated from the model show, while the largest AUC values by variables that are added individually (Phillips et al., 2006). In order to analyze the prediction accuracy, Maxent automatically divides our mosquito sample information into training and test sets. Both threshold-independent and threshold-dependent methods are used to evaluate how accurate the model predictions are (Phillips et al., 2006). The threshold independent approach uses external omission rate (the percentage of test sites that fall in pixels that are not suitable for the species) and proportional projected area (the percentage of all pixels that are favorable for the species) to evaluate the model prediction. Despite the fact that independent threshold assessments utilize the recipient's operational features to measure the area under the curve.

Receiver Operating Characteristics Curves (ROC)

Maxent's performance is indicated by the area under the curve (AUC) value, which is typically the unit square section of the area with a value between 0 and 1. Using this receiver operating feature approach, Maxent predicts the distribution according to species (Fawcett, 2006). This curve shows how well a classifier works when a threshold parameter affects its output. This graph shows the ratio of true and false positive values for each level. The classifier has classifies a fraction "x" of negative examples (grids without occurrence localities) as positive and a proportion "y" of positive cases as positive shown by some location on the graph (Phillips, 2004). The points on the graph are joined to create a curve. However, the area under the curve (AUC) shows how likely it is

that the classifier will correctly arrange the two points (Phillips, 2004). An AUC value of less than one indicates a good classifier. (Wiley and others, 2003). Threshold unbiased measurements show sensitivity and specificity on a graph. Sensitivity is the accurately true positive predicted sites for a species, while selectivity is the real, exactly determined negative sites (Fielding and Bell, 1997; Phillips et al., 2006; Phillips and Dudik, 2008). The AUC value can be computed by the differential capacity of the model. 0.5 to 0.7, 0.7 to 0.9, or > 0.9 and 1.0 are AUC values of the model's low, fair, high, and perfect discriminative power correspondingly (Brooker et al., 2001; Phillips et al., 2006; Phillips and Dudik, 2008).

Results and Discussion

In species distribution modeling (SDM), visual tools such as response curves and the jackknife test play a vital role in exploring how environmental factors shape the presence or spread of a species. Response curves visually depict the influence of individual environmental variables on species predictions, helping researchers understand which factors are most critical and which have limited impact. Interpreting these curves, however, can be challenging when variables are closely related, as overlapping effects may obscure clear patterns. Alongside these, the jackknife test serves as a powerful method to evaluate the model's reliability by systematically excluding variables or data points to observe changes in prediction accuracy. This process highlights potential overfitting and helps determine how well the model can handle unseen data. Together, these methods provide a clearer picture of ecological relationships and support efforts in species conservation and environmental planning.

Results

Mosquito distribution in Faisalabad

Larvae of *Aedes* were collected from different sites in Faisalabad city. Table discusses their websites in more detail.

Analysis of omission/commission

The forecasted area and exclusion ratio are shown against the progressive approach in the following figure. The original presence records and, if test data is used, the test records as well are used to evaluate the exclusion ratio. For this reason the specification of the progressive approach, the exclusion ratio must be around the predicted omission.

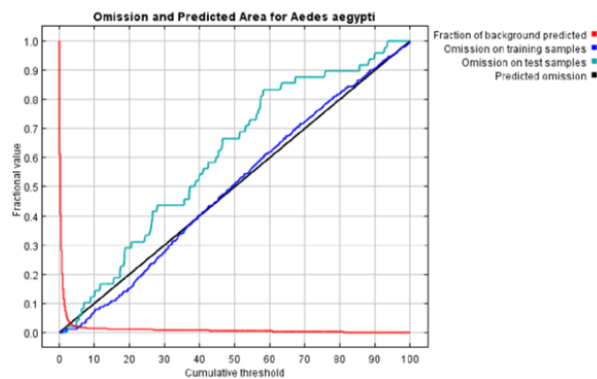


Figure 4. Omission and predicted are for *Aedes aegypti*

In the following figure the receiver operating characteristic (ROC) curve for the similar data is shown. Keep in mind that specificity is determined by expected area rather than actual commission; see the Phillips, Anderson, and Schapire study area to understand about its, on the support page. This suggests that the highest AUC that may be attained is less than 1. The maximum test AUC that could be achieved if test data were taken from the Maxent distribution itself would be 0.986 instead of 1, even if the test AUC may be greater.

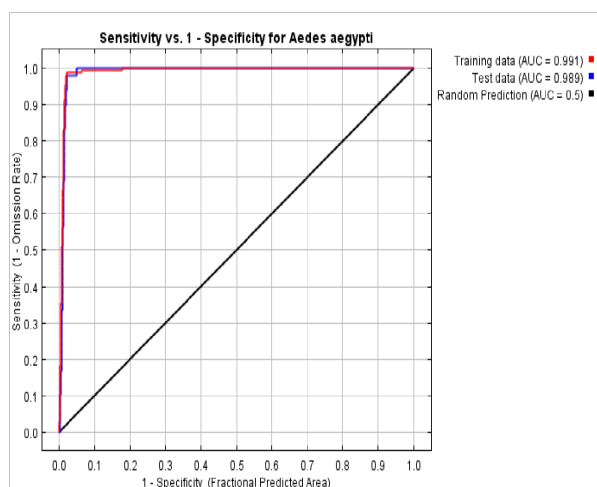


Figure 5. Sensitivity vs. 1-Specificity for *Aedes aegypti*

Below is a list of some specific criteria along with the corresponding exclusion ratios. When test information is provided, if there are less than 25 test samples, binomial chances are computed appropriately; if not, a conventional method to the binomial is used. The null hypothesis, which states that test sites can be expected no more accurately than by a random prediction with the same fractional anticipated area, is supported by these one-sided p-values. The total of the following is reduced by the "Balance" threshold: Six * fractional predicted area + .04 * cumulative threshold + 1.6 * training omission rate.

Table 2. Maxent modeling analysis of *Aedes aegypti* and its output.

Cumulative threshold	Logistic threshold	Description	Fractional Predicted area	Training omission rate	Test omission rate	P-value
1.000	0.001	Fixed cumulative value 1	0.169	0.007	0.000	1.42E-53
5.000	0.085	Fixed cumulative value 5	0.021	0.014	0.042	0E0
10.000	0.313	Fixed cumulative value 10	0.016	0.075	0.125	0E0
0.967	0.001	Minimum training presence	0.176	0.000	0.000	3.849E-51
13.773	0.430	10 percentile training presence	0.014	0.095	0.167	0E0
5.470	0.106	Equal training sensitivity and specificity	0.020	0.020	0.042	0E0
5.004	0.085	Maximum training sensitivity plus specificity	0.021	0.014	0.042	0E0
4.855	0.079	Equal test sensitivity and specificity	0.022	0.014	0.021	0E0
4.855	0.079	Maximum test sensitivity plus specificity	0.022	0.014	0.021	0E0
1.997	0.004	Balance training omission, predicted area and threshold value	0.063	0.007	0.000	0E0
4.256	0.057	Equate entropy of thresholded and original distributions	0.024	0.014	0.021	0E0

Response curves

In these graphs the effect of every environmental variable on the Maxent forecast is shown. When each environmental factor is changed while maintaining a fixed mean sampling value for all other environmental variables, the graphs illustrate how logistical prediction varies. It perhaps be demanding to clarify the curves if the variables that you employ are closely related since the model could rely on the correlations in ways that are not shown in the curves. In other words, the curves demonstrate the minimal effect of changing a single variable, even when the model could advantageous from changing groups of variables collectively.

Molecular Docking Analysis of DPP-4 Enzyme with Polyphenol Compounds

Similar to AG enzyme, all the selected polyphenol compounds and sitagliptin were successfully docked at the targeted binding sites of the DPP-4 enzyme. The results of the molecular docking for each combination that formed a complex with DPP-4 enzyme were summarized in Table 3 with their corresponding binding energy values. In Table 4, it stated the results of the hydrogen bond formation and the interacting residues of DPP-4 with each polyphenol compound and sitagliptin.

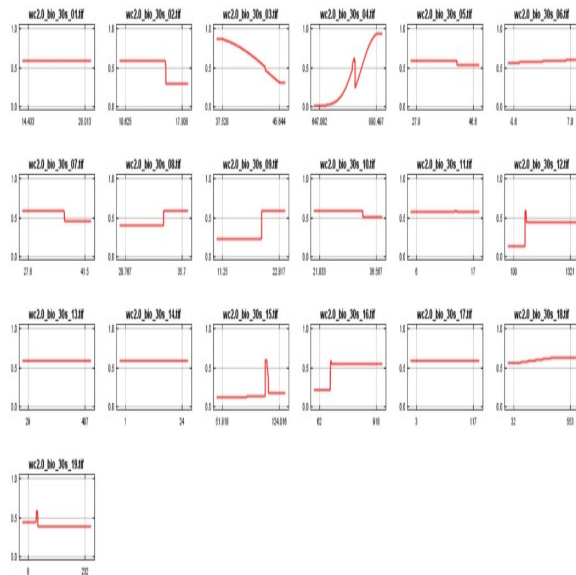


Figure 6. Responsive curves in accordance to each environmental variable.

Every following curves represents a different model, compared to the marginal response curves above, i.e.,

Table 3. Estimate for the Relative Contributions of Ecological Factors.

Variable	Percent contribution	Permutation importance
wc2.0_bio_30s_15.tif	19.9	5.7
wc2.0_bio_30s_16.tif	18.4	12.6
wc2.0_bio_30s_12.tif	11.3	22.1
wc2.0_bio_30s_19.tif	9.8	0
wc2.0_bio_30s_09.tif	9.6	24.1
wc2.0_bio_30s_08.tif	8.6	0.7
wc2.0_bio_30s_03.tif	7.1	7.1
wc2.0_bio_30s_02.tif	5.1	7.2
wc2.0_bio_30s_07.tif	4.6	0
wc2.0_bio_30s_10.tif	3.8	0
wc2.0_bio_30s_04.tif	1.7	20.4
wc2.0_bio_30s_05.tif	0.1	0
wc2.0_bio_30s_18.tif	0	0
wc2.0_bio_30s_13.tif	0	0
wc2.0_bio_30s_11.tif	0	0.2
wc2.0_bio_30s_06.tif	0	0
wc2.0_bio_30s_17.tif	0	0
wc2.0_bio_30s_14.tif	0	0
wc2.0_bio_30s_01.tif	0	0

The Jackknife test of constant relevancy findings are shown in the following image. Since it has the most benefit when utilized alone, the ecological factor wc2.0_bio_30s_12.tif seems to contain the most valuable information. Since Wc2.0_bio_30s_15.tif is the component of the environment when deleted, decreases the gain the most, it looks like to have the most of data that is absent in the other variables.

a Maxent model developed using just related variable. The graphs show the weakness of forecasted appropriateness on the variable of choice and weaknesses resulting from association between other factors and the variable of choice. If there are strong relationships between them, the variables may be simple to comprehend.

Analysis of variable contributions:

The given table presents predictions about comparative contribution made by environmental factors to the Maxent model. In each training process iteration, the initial estimate is computed by adding the addition of the established gain to the related variable's contributions, or its removal in the event that fluctuate the true worth of lambda is negative. The learning presence and background data of every single environmental component are permuted at random for the second evaluation. The training AUC drop resulting from reevaluating the prediction model on the permuted data is shown in the table, adjusted to percentages. When the variables that predict are connected, variable contributors should be carefully evaluated, much as the variable jackknife



Figure 6. Jackknife of regularised training gain for *Aedes aegypti*.

In the following picture the identical Jackknife test is shown, but test gain is used in place of training gain. Keep in mind that test findings may cause us to change our minds about which aspects are most crucial.

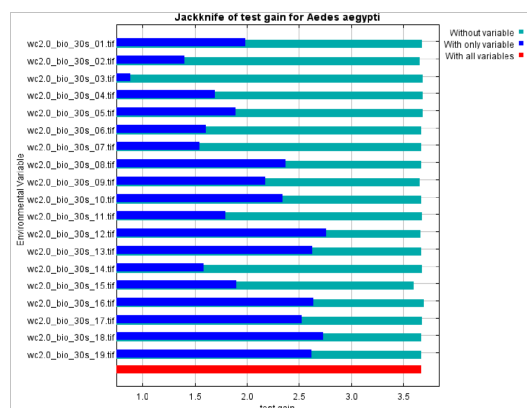


Figure 7. Jackknife of test gain for *Aedes aegypti*

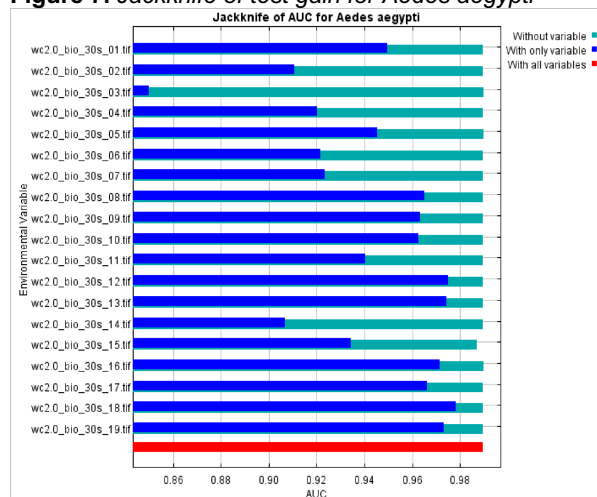


Figure 8. Jackknife of AUC gain for *Aedes aegypti*

Discussion

The graphics that are displayed shed light on a critical aspect of species distribution modeling (SDM) by emphasizing reaction curves and the understanding of Maxent models. These visualization approaches are essential for comprehending how environmental conditions impact the projected distribution of species.

Response curves must be understood in order to comprehend the relationship between environmental conditions and species dispersion forecasts. By examining these response curves, we can ascertain the extent to which changes in a particular environmental variable impact a species' predicted distribution. Interpreting response curves is crucial to understanding the ecological interactions between species and their environments. If a response curve for a specific environmental variable is generally flat through most of range of values, it may not have a powerful effect on distribution of species. In particular, the presence of significant correlations between environmental variables also entangled the interpretation. In this scenario the model may manipulate connections between factors in such ways that response curves do not readily show. Contrastingly to the marginal reaction curves which were addressed previously, these graphs describe how the selected variables expected to appropriately affects selected variables along with the other factors

to produce dependencies. The graphs is in depth visually describe how the change of environmental circumstances impact the expected expansion of species. These graphs enable the researchers to understand the pattern of ecological processes in controlling specie distribution.

Concluding that response curves along with Maxent model response curves are the most crucial tools used by researchers to develop relationship between environmental factors on species distribution, how they understand species distribution. This research graphs are a quick way to understand the ecological dynamics which are effecting species distribution and steps can be taken to conserve species and managing the environmental conditions to the extent they can.

The jackknife test is as essential tool as the previously described response curve and Maxent model response are used for the SDM (species distribution modeling). Jackknife test is a resampling method used to determine the statistical estimator's variance and bias. After understanding model on all the other data points, this cross-validation is called leave-one-out, and it is used to test the model on excluded data point. In this process the test is repeated for each data point in the dataset. The jackknife testis used to draw graphs that depicts the Maxent models performance and it's stability in that particular research. By comparing actual and expected values of every missing data point research can evaluate the accuracy of models' predictions. The jackknife test helps in identifying over fitting model by evaluating the model's performance on imagined data points. If prediction across all the data points came out consistent then it model appears to be stable and suitable for handling unknown data. Over fitting, are the model's where new data effect the model's ability to detect its target. If the jackknife test predictions are unlike from the training data predictions than this is over fitting model.

Conclusions

The study provides valuable information about the geographic dispersal region for the important mosquito species that act as vector represent a major health risk. To prevent the spread of illness, selected vector management measures are necessary. The predictive models developed in this work can help direct efforts to manage vectors in the regionof vector mosquitoes in the urban regions of Pakistan especially Faisalabad, using GIS techniques. Since the area is main suitable breeding.

There are various restrictions on this study. Because of the district's terrain in some places, mosquito captures from the allocated locations were insufficient for field validations of these models. Finding high-risk areas close to water sources and mosquito populations was one way to deal with this problem. Positive results were occasionally achieved, even though these efforts were not always successful. In particular, accessibility was a major obstacle to the collection of *Ae. aegypti* from high-risk areas; just two sites produced positive results, and the rest sites

produced negative results because of a lack of water or logistical challenges in getting to them. However, our findings are supported by the two positive locations in high-risk zones, which validate our model predictions.

In Faisalabad, the region of Punjab in Pakistan, there is spread of diseases affecting its urban areas; these diseases are carried by mosquitoes, so some targeted vector control measures are needed to put in that place. Moreover, it is needed to analyze the directional proficiency of mosquito species at different places. So in order to check the vector potential and dispersion in metropolis of Faisalabad, Punjab, Pakistan, there is need of more larvae collection of mosquito. There are some strategies we can adopt to control this disease spreading by vector by different controlling strategies that includes use of pesticide spray, source reduction of larvae, reduction of mosquito breeding grounds, control of mosquito population, improvement of sanitary conditions, improvement of waste management infrastructure, implementation of IVM (integrated vector management) as it contains a number of control mechanisms, public awareness about the causative agents of diseases, carry out a range of mosquito control programs with the help of environmental departments etc.

Author Contributions

S.S.: Writing – original draft, Visualization, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. M.R. and M.N.: Writing – review & editing, Visualization, Formal analysis, Conceptualization. H.C. and N.H.B.: Writing – review & editing, Validation, Supervision, Resources, Project administration, Funding acquisition. Wei Luo: Writing – review & editing, Conceptualization. N.R.: Writing – review & editing, Conceptualization. R.Y.: Writing – review & editing.

Conflicts of Interest

The author(s) declare(s) that there is no conflict of interest regarding the publication of this paper.

Acknowledgment

We are thankful for the support of this study to Riphah International University, Faisalabad Campus.

Data Availability

Data will be made available on request.

References

1. Abdel-Dayem, M. S., Annajar, B. B., Hanafi, H. A., & Obenauer, P. J. (2012). The potential distribution of *Phlebotomus papatasi* (Diptera: Psychodidae) in Libya based on ecological niche model. *Journal of Medical Entomology*, 49(3), 739-745.
2. Agarwal, S. A., Sikarwar, S. S., & Sukumaran, D. (2012). Application of remote sensing & GIS in risk area assessment for mosquito borne diseases - A case study in a part of Gwalior City (M. P.). *International Journal of Advanced Technology & Engineering Research*, 2(1), 1-4.
3. Ahmad, H., Ali, A., Fatima, S. H., Zaidi, F., Khisroon, M., Rasheed, S. B., Ullah, I., Ullah, S., & Shakir, M. (2020). Spatial modeling of dengue prevalence and kriging prediction of dengue outbreak in Khyber Pakhtunkhwa (Pakistan) using presence only data.
4. *Stochastic Environmental Research and Risk Assessment*, 34(7), 1023-1036.
5. Ahmad, R., Ali, W. N. W. M., Nor, Z. M., Ismail, Z., Hadi, A. A., Ibrahim, M. N., & Lim, L. H. (2011). Mapping of mosquito breeding sites in malaria endemic areas in Pos Lenjang, Kuala Lipis, Pahang, Malaysia. *Malaria Journal*, 10, 1-12.
6. Al Ahmed, A. M., Al Kuriji, M. A., Kheir, S. M., Al Sogoor, D. A., & Salama, H. A. (2010). Distribution and seasonal abundance of mosquitoes (Diptera: Culicidae) in the Najran Region, Saudi Arabia. *Stud Dipterol*, 17, 13-27.
7. Alahmad, A. M., Sallam, M. F., Khuriji, M. A., Kheir, S. M., & Azari-Hamidian, S. (2011). Checklist and pictorial key to fourth-instar larvae of mosquitoes (Diptera: Culicidae) of Saudi Arabia. *Journal of Medical Entomology*, 48(4), 717-737.
8. Alahmed, A. M., Kheir, S. M., Kuriji, M. A., & Sallam, M. F. (2011). Breeding habitats characterization of *Anopheles* mosquito (Diptera: Culicidae) in Najran Province, Saudi Arabia. *Journal of the Egyptian Society of Parasitology*, 41(2), 275-288.
9. Alahmed, A. M., Naeem, M., Kheir, S. M., & Sallam, M. F. (2015). Ecological distribution modeling of two malaria mosquito vectors using geographical information system in Al-Baha Province, Kingdom of Saudi Arabia. *Pakistan Journal of Zoology*, 47(6), 1797-1806.
10. Alahmed, A. M., Shaalan, E. A., Aboul-Soud, M. A. M., Tripet, F., & Al-Khedhairi, A. A. (2011). Mosquito vectors survey in the Al-Ahsaa district of eastern Saudi Arabia. *Journal of Insect Science*, 11(3), 335-370.
11. Ali, S. A., & Ahmad, A. (2019). Mapping of mosquito-borne diseases in Kolkata Municipal Corporation using GIS and AHP based decision making approach. *Spatial Information Research*, 27(3), 351-372.
12. Al-Thukair, A., Jemal, Y., & Nzila, A. (2022). Influence of climatic factors on the abundance and profusion of mosquitoes in Eastern Province, Saudi Arabia. In *Mosquito ResearchRecent Advances in Pathogen Interactions, Immunity, and Vector Control Strategies*. IntechOpen, 23(5), 103-115.
13. Ammar, M., Moaaz, M., Yue, C., Fang, Y., Zhang, Y., Shen, S., & Deng, F. (2025). Emerging Arboviral Diseases in Pakistan: Epidemiology and Public Health Implications. *Viruses*, 17(2), 232.
14. Amer, A., & Mehlhorn, H. (2006). Repellency effect of forty-one essential oils against *Aedes*, *Anopheles*, and *Culex* mosquitoes. *Parasitology research*, 99, 478-490.
15. Anderson, R. P., & Martínez-Meyer, E. (2004). Modeling species' geographic distributions for preliminary conservation assessments: an implementation with the spiny pocket mice (Heteromys) of Ecuador. *Biological Conservation*, 116, 167-179.
16. Anderson, R. P., Gómez-Laverde, M., & Peterson, A. T. (2002). Geographical distributions of spiny pocket mice in South America: insights from predictive models. *Global Ecology and Biogeography*, 11, 131-141.
17. Andreadis, T. G., Anderson, J. F., Vossbrinck, C. R., & Main, A. J. (2004). Epidemiology of West Nile virus in Connecticut: a five-year analysis of mosquito data 1999–2003. *Vector-Borne & Zoonotic Diseases*, 4(4),

- 360-378.
19. Anyamba, A., & Tucker, C. J. (2005). Analysis of Sahelian vegetation dynamics using NOAAAVHRR NDVI data from 1981–2003. *Journal of Arid Environments*, 63, 569–614.
 20. Bagavan, A., Rahuman, A. A., Kamaraj, C., & Geetha, K. (2008). Larvicidal activity of saponin from *Achyranthes aspera* against *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitology research*, 103, 223–229.
 21. Barbet-Massin, M., Rome, Q., Villemant, C., & Courchamp, F. (2018). Can species distribution models really predict the expansion of invasive species? *PLoS ONE*, 13(3), 1–14.
 22. Beck, L. R., Rodriguez, M. H., Dister, S. W., et al. (1994). Remote sensing as a landscape epidemiologic tool to identify villages at high risk for malaria transmission. *American Journal of Tropical Medicine and Hygiene*, 51(3), 271–280.
 23. Beck, L. R., Rodriguez, M. H., Dister, S. W., et al. (1997). Assessment of a remote sensing–based model for predicting malaria transmission risk in villages of Chiapas, Mexico. *American Journal of Tropical Medicine and Hygiene*, 56(1), 99–106.
 24. Booman, M., Durrheim, D. N., Grange, K. La., Martin, C., Mabuza, A. M., Zitha, A., Mbokazi, F. M., Fraser, C., & Sharp, B. L. (2000). Using a geographical information system to plan a malaria control programme in South Africa. *Bulletin of the World Health Organization*, 78, 1438–1444.
 25. Brooker, S., Hay, S. I., Issae, W., Hall, A., Kihamia, C. M., Lwambo, N. J., Wint, W., Rogers, D. J., & Bundy, D. A. (2001). Predicting the distribution of urinary schistosomiasis in Tanzania using satellite sensor data. *Tropical Medicine & International Health*, 6(12), 998–1007.
 27. Brownstein, J. S., Holford, T. R., Fish, D. (2004). Enhancing West Nile virus surveillance, United States. *Emerging Infectious Diseases*, 10(6), 1129–1133.
 28. Brownstein, J. S., Rosen, H., Purdy, D., Miller, J. R., Merlino, M., Mostashari, F., & Fish, D. (2002). Spatial analysis of West Nile virus: rapid risk assessment of an introduced vector-borne zoonosis. *Vector Borne and Zoonotic Diseases*, 2(3), 157–164.
 29. Carter, R., Mendis, K. N., & Roberts, D. (2000). Spatial targeting of interventions against malaria. *Bulletin of the World Health Organization*, 78, 1401–1411.
 30. Chinery, W. A. (1984). Effects of ecological changes on the malaria vectors *Anopheles funestus* and the *Anopheles gambiae* complex of mosquitoes in Accra, Ghana. *The American Journal of Tropical Medicine and Hygiene*, 87, 75–81.
 31. Cleckner, H. L., Allen, T. R., & Scott Bellows, A. (2011). Remote sensing and modeling of mosquito abundance and habitats in Coastal Virginia, USA. *Remote Sensing*, 3(12), 2663–2681.
 32. Cocu, N., Conrad, K., Harrington, R., & Rounsevell, M. D. A. (2005). Analysis of spatial patterns at a geographical scale over north-western Europe from point-referenced aphid count data. *Bulletin of entomological research*, 95(1), 47–56.
 33. Coetzee, M., Craig, M., & Le Sueur, D. (2000). Distribution of African malaria mosquitoes belonging to the *Anopheles gambiae* complex. *Parasitology Today*, 16(2), 74–77.
 34. Cohuet, A., Harris, C., Robert, V., & Fontenille, D. (2010). Evolutionary forces on *Anopheles*:
 35. What makes a malaria vector? *Trends in Parasitology*, 26(3), 130–136.
 36. Conley, A. K., Fuller, D. O., Haddad, N., Hassan, A. N., Gad, A. M., & Beier, J. C. (2014). Modeling the distribution of the West Nile and Rift Valley Fever vector *Culex pipiens* in arid and semi-arid regions of the Middle East North Africa. *Parasites & Vectors*, 7, 289.
 37. Craig, M. H., Snow, R. W., & Le Sueur, D. (1999). A climate-based distribution model of malaria transmission in sub-Saharan Africa. *Parasitology Today*, 15(3), 105–111.
 38. Cross, E. R., Newcomb, W. W., & Tucker, C. J. (1996). Use of weather data and remote sensing to predict the geographic and seasonal distribution of *Phlebotomus papatasi* in southwest Asia. *The American journal of tropical medicine and hygiene*, 54(5), 530–536.
 39. Dambach, P., Machault, V., Lacaux, J. P., Vignolles, C., Sie, A., & Sauerborn, R. (2012). Utilization of combined remote sensing techniques to detect environmental variables influencing malaria vector densities in rural West Africa. *International Journal of Health Geographics*, 11, 8.
 40. DeGroot, J. P., Sugumaran, R., Brend, S. M., Tucker, B. J., & Bartholomay, L. C. (2008). Landscape, demographic, entomological, and climatic associations with human disease incidence of West Nile virus in the state of Iowa, USA. *International Journal of Health Geographics*, 7, 19.
 41. Dister, S. W., Fish, D., Bros, S. M., Frank, D. H., & Wood, B. L. (1997). Landscape characterization of peridomestic risk for Lyme disease using satellite imagery. *The American journal of tropical medicine and hygiene*, 57(6), 687–692.
 42. Diuk-Wasser, M. A., Bogayoko, M., Sogoba, N., et al. (2004). Mapping rice field *anopheline* breeding habitats in Mali, West Africa, using Landsat ETM sensor data. *International Journal of Remote Sensing*, 25(2), 359–376.
 43. Diuk-Wasser, M. A., Brown, H. E., Andreadis, T. G., & Fish, D. (2006). Modeling the spatial distribution vectors for West Nile virus in Connecticut, USA. *EcoHealth*, 6(3), 1–13.
 44. Dutta, P., Bhattacharyya, D. R., Sharma, C. K., Khan, S. A., & Mahanta, J. (1998). Distribution of potential dengue vectors in major townships along the national highway and trunk roads of north east India. *Southeast Asian Journal of Tropical Medicine and Public Health*, 29, 173–176.
 45. Eisen, L., & Eisen, R. J. (2011). Using geographic information systems and decision support systems for the prediction, prevention, and control of vector-borne diseases. *Annual review of entomology*, 56, 41–61.
 46. Elfadil, A. A., Hasab-Allah, K. A., & Dafa-Allah, O. M. (2006). Factors associated with rift valley fever in south-west Saudi Arabia. *Revue Scientifique et Technique (International Office of Epizootics)*, 25(3), 1137–1145.
 47. Elith, J., H. Graham, C., P. Anderson, R., Dudík, M., Ferrier, S., Guisan, A., & E.
 48. Zimmermann, N. (2006). Novel methods improve prediction of species' distributions from occurrence data. *Ecography*, 29(2), 129–151.
 49. Elnaiem, D. E., Schorschew, J., et al. (2003). Risk mapping of visceral leishmaniasis: The role of local variation in rainfall and altitude on the presence and incidence of kala-azar in eastern Sudan. *American Journal of Tropical Medicine and Hygiene*, 68(1), 10–17.
 50. Fawcett, T. (2006). An introduction to ROC analysis. *Pattern Recognition Letters*, 27, 861–874.
 51. Fielding, A. H., & Bell, J. F. (1997). A review of methods

- for the assessment of prediction errors in conservation presence/absence models. *Environmental Conservation*, 24, 38-49.
52. Foley, D. H., Klein, T. A., Kim, H. C., Brown, T., Wilkerson, R. C., & Rueda, L. M. (2010). Validation of ecological niche models for potential malaria vectors in the Republic of Korea. *Journal of the American Mosquito Control Association*, 26(2), 210-213.
 53. Fossog, B., Ayala, D., Acevedo, P., Kengne, P., Ngomo Abeso Mebuy, I., Makanga, B., Magnus, J., Awono-Ambene, P., Njiokou, F., Pombi, M., Antonio-Nkondjio, C., Paupy, C., Besansky, N. J., & Costantini, C. (2015). Habitat segregation and ecological character displacement in cryptic African malaria mosquitoes. *Evolutionary Applications*, 8(4), 326-345.
 54. Gakhar, S. K., Sharma, R., & Sharma, A. (2013). Population genetic structure of malaria vector *Anopheles stephensi* Liston (Diptera: Culicidae). *Indian Journal of Experimental Biology*, 51, 273-279.
 55. Garcia-Rejon, J. E., Blitvich, B. J., Farfan-Ale, J. A., Loroño-Pino, M. A., Chi Chim, W. A., Flores-Flores, L. F., & Beaty, B. J. (2008). Dengue virus infected *Aedes aegypti* in the home environment. *American Journal of Tropical Medicine and Hygiene*, 79, 940-950.
 - Ghebreyesus, T. A., Haile, M., Getachew, A., Alemayehu, T., Witten, K. H., Medhin, A., & Byass, P. (1998). Pilot studies on the possible effects on malaria of small-scale irrigation dams in Tigray regional state, Ethiopia. *Journal of Public Health Medicine*, 20(2), 238-240.
 56. Glick, J. I. (1992). Illustrated key to the female *Anopheles* of southwestern Asia and Egypt (Diptera: Culicidae). *Mosquito Systematics Journal*, 24, 125-153.
 57. Goodchild, M. F. (1992). Geographical Information Science. *International Journal of Geographical Information Systems*, 6(1), 31-45.
 58. Goodchild, M. F. (2009). Geographic information systems and science: Today and tomorrow. *Annals of GIS*, 15(1), 3-9.
 59. Gubler, D. J. (1998). Dengue and dengue hemorrhagic fever. *Clinical Microbiology Reviews*, 11(3), 480-496.
 60. Guerra, M. A., Walker, E. D., & Kitron, U. (2001). Canine surveillance system for Lyme borreliosis in Wisconsin and northern Illinois: geographic distribution and risk factor analysis. *American Journal of Tropical Medicine and Hygiene*, 65(6), 546-552.
 61. Guerra, M., Walker, E., Jones, C., et al. (2002). Predicting the risk of Lyme disease: habitat suitability for Ixodes scapularis in the north central United States. *Emerging Infectious Diseases*, 8(3), 289-297.
 62. Guisan, A., & Zimmerman, N. E. (2000). Predictive habitat distribution models in ecology. *Ecological Modelling*, 135(2-3), 147-186.
 63. Gurgel-Gonçalves, R., Galvão, C., Costa, J., & Peterson, A. T. (2012). Geographic distribution of Chagas disease vectors in Brazil based on ecological niche modeling. *Journal of tropical medicine*, 2012(1), 705326.
 64. Harbach, R. E. (1985). Pictorial keys to the genera of mosquitoes, subgenera of *Culex*, and the species of *Culex* (*Culex*) occurring in southwestern Asia and Egypt, with a note on the subgeneric placement of *Culex deserticola* (Diptera: Culicidae). *Mosquito Systematics*, 17, 83-107.
 65. Harbach, R. E. (1988). The mosquitoes of the subgenus *Culex* in southwestern Asia and Egypt (Diptera: Culicidae). *Contributions of American Entomological Institute*, 24, 1-240.
 66. Hay, S. I., Omumbo, J. A., Craig, M. H., & Snow, R. W. (2000). Earth observation, geographic information systems and Plasmodium falciparum malaria in sub-Saharan Africa. *Advances in Parasitology*, 47, 173-215.
 67. Hay, S. I., Packer, M. J., & Rogers, D. J. (1997). The impact of remote sensing on the study and control of invertebrate intermediate hosts and vectors for disease. *International Journal of Remote Sensing*, 18, 2899-2930.
 68. Hay, S. I., Sinka, M. E., Okara, R. M., Kabaria, C. W., Mbithi, P. M., Tago, C. C., & Godfray, H. C. J. (2010). Developing global maps of the dominant *Anopheles* vectors of human malaria. *PLoS medicine*, 7(2).
 69. Hernandez, P. A., Graham, C. H., Master, L. L., & Albert, D. L. (2006). The effect of sample size and species characteristics on performance of different species distribution modeling methods. *Ecography*, 29, 773-785.
 70. Holeva-Eklund, W. M., Young, S. J., Will, J., Busser, N., Townsend, J., & Hepp, C. M. (2022). Species distribution modeling of *Aedes aegypti* in Maricopa County, Arizona from 2014 to 2020. *Frontiers in Environmental Science*, 10.
 71. Huang, J., Walker, E. D., Otienoburu, P. E., Amimo, F., Vulule, J., & Miller, J. R. (2006). Laboratory tests of oviposition by the African malaria mosquito, *Anopheles gambiae*, on dark soil as influenced by presence or absence of vegetation. *Malaria Journal*, 5, 88.
 72. Hunt, R. H., Fuseini, G., Knowles, S., Stiles-Ocran, J., Verster, R., Kaiser, M. L., Choi, K. S., Koekemoer, L. L., & Coetzee, M. (2011). Insecticide resistance in malaria vector mosquitoes at four localities in Ghana, West Africa. *Parasites & Vectors*, 4, 107.
 73. Hunter, J. M., Rey, L., & Scott, D. (1982). Man-made lakes and man-made diseases. Towards a policy resolution. *Social Science & Medicine*, 16, 1127-1145.
 74. Hutchinson, G. E. (1957). Concluding remarks. *Cold Spring Harbor Symposia on Quantitative Biology*, 22, 415-427.
 75. Jansen, C. C., & Beebe, N. W. (2010). The dengue vector *Aedes aegypti*: what comes next? *Microbes and Infection*, 12(4), 272-279.
 76. Jaynes, E. T. (1957). Information theory and statistical mechanics. *Physical Review*, 106, 620-630.
 77. Jeffery, J. A., Ryan, P. A., Lyons, S. A., Thomas, P. T., & Kay, B. H. (2002). Spatial distribution of vectors of Ross River virus and Barmah Forest virus on Russell Island, Moreton Bay, Queensland. *Australian Journal of Entomology*, 41(4), 329-338.
 78. Jemal, Y., & Al-Thukair, A. A. (2018). Combining GIS application and climatic factors for mosquito control in Eastern Province, Saudi Arabia. *Saudi Journal of Biological Sciences*, 25(8), 1593-1602.
 79. Kazmi, J. H., & Pandit, K. (2001). Disease and dislocation: The impact of refugee movements on the geography of malaria in NWFP, Pakistan. *Social Science & Medicine*, 52(7), 1043-1055.
 80. Khaliq, A., Chaudhry, M. N., Sajid, M. A., Ashraf, U., Aleem, R., & Shahid, S. (2021). GIS based mapping and spatial distribution of tuberculosis in Punjab, Pakistan. *Epidemiology Science*, 11(402), 1-6.
 81. Kheir, S. M., Alahmed, A. M., Al Kuriji, M. A., & Al Zubayani, S. F. (2010). Distribution and seasonal activity of mosquito in Al Madinah Al Munwarrah, Saudi Arabia. *Journal of the Egyptian Society of Parasitology*, 40(1), 215-227.
 82. Kija, B., Mwera, C., Mwita, M., & Fumagwa, R. (2013). Prediction of suitable habitat for potential invasive plant

- species *Parthenium hysterophorus* in Tanzania: A short communication. *International Journal of Ecosystem*, 3, 82-89.
84. Kolimenakis, A., Heinz, S., Wilson, M. L., Winkler, V., Yakob, L., Michaelakis, A., ... & Horstick, O. (2021). The role of urbanisation in the spread of *Aedes* mosquitoes and the diseases they transmit—A systematic review. *PLoS neglected tropical diseases*, 15(9), e0009631.
 85. Kitron, U., & Spielman, A. (1989). Suppression of transmission of malaria through source reduction: Antianopheline measures applied in Israel, the United States, and Italy. *Review of Infectious Diseases*, 11, 391-406.
 86. Kitron, U., Okeno, L. H., Hungerford, L. L., et al. (1996). Spatial analysis of the distribution of tsetse flies in the Lambwe Valley, Kenya, using Landsat TM satellite imagery and GIS. *Journal of Animal Ecology*, 65(3), 371-380.
 87. Kitron, U., Pener, H., Costin, C., Orshan, L., Greenberg, Z., & Shalom, U. (1994). Geographic information system in malaria surveillance: mosquito breeding and imported cases in Israel, 1992. *American Journal of Tropical Medicine and Hygiene*, 50, 550-556.
 88. Kolimenakis, A., Heinz, S., Wilson, M. L., Winkler, V., Yakob, L., Michaelakis, A., ... & Horstick, O. (2021). The role of urbanisation in the spread of *Aedes* mosquitoes and the diseases they transmit—A systematic review. *PLoS neglected tropical diseases*, 15(9), e0009631.
 89. Kolivras, K. N. (2006). Mosquito habitat and dengue risk potential in Hawaii: A conceptual framework and GIS application. *Professional Geographer*, 58(2), 139-154.
 90. Kolivras, K. N. (2010). Changes in dengue risk potential in Hawaii, USA, due to climate variability and change. *Climate Research*, 42(1), 1-11. <https://doi.org/10.3354/cr00861>
 91. Kulkarni, M. A., Desrochers, R. E., & Kerr, J. T. (2010). High resolution niche models of malaria vectors in northern Tanzania: a new capacity to predict malaria risk? *PLoS One*, 5(2).
 92. Larson, S. R., DeGroot, J. P., Bartholomay, L. C., & Sugumaran, R. (2010). Ecological niche modeling of potential West Nile virus vector mosquito species in Iowa. *Journal of Insect Science*, 10, 110.
 93. Lin, T. H., & Lu, L. C. (1995). Population fluctuation of *Culex tritaeniorhynchus* in Taiwan.
 94. *Chinese Journal of Entomology*, 15, 1-9.
 95. Linthicum, K. J., Anyamba, A., Tucker, C. J., Kelley, P. W., Myers, M. F., & Peters, C. J. (1999). Climate and satellite indicators to forecast Rift Valley fever epidemics in Kenya. *Science*, 285, 397-400.
 96. Losos, J. B., Leal, M., Glor, R. E., de Queiroz, K., Hertz, P. E., Schettino, L. R., Lara, A. C. (2003). Niche lability in the evolution of a Caribbean lizard community. *Nature*, 424(6948), 542-545.
 97. Luckhart, S., Vodovotz, Y., Cui, L., & Rosenberg, R. (1998). The mosquito *Anopheles stephensi* limits malaria parasite development with inducible synthesis of nitric oxide.
 98. *Proceedings of the National Academy of Sciences of the USA*, 95, 5700-5705.
 99. Mackenzie, J. S., et al. (1996). Dengue in Australia. *Journal of Medical Microbiology*, 45, 159-161.
 100. Madani, T. A., Al-Mazrou, Y. Y., Al-Jeffri, M. H., Mishkhas, A. A., Al-Rabeah, A. M., Turkistani, A. M., Al-Sayed, M. O., Abodahish, A. A., Khan, A. S., Ksiazek, T. G., & Shobokshi, O. (2003). Rift Valley Fever epidemic in Saudi Arabia: Epidemiological, clinical, and laboratory characteristics. *Clinical Infectious Diseases*, 37(8), 1084-1092.
 101. Mahabir, R. S., Severson, D. W., & Chadee, D. D. (2012). Impact of road networks on the distribution of dengue fever cases in Trinidad, West Indies. *Acta Tropica*, 123, 178-183.
 102. Malik, M. A., Sajid, M. S., Iqbal, Z., & Saqib, M. (2023). Association of climatic determinants with the magnitude of *Aedes aegypti* in selected agro-geoclimatic zones of Punjab, Pakistan, 6(3), 133-141.
 103. Masuoka, P., Klein, T. A., Kim, H. C., Claborn, D. M., Achee, N., Andre, R., & Grieco, J. (2010). Modeling the distribution of *Culex tritaeniorhynchus* to predict Japanese encephalitis distribution in the Republic of Korea. *Geospatial Health*, 5(1), 45-57.
 104. Mathew, N., Anitha, M. G., Bala, T. S. L., Sivakumar, S. M., Narmadha, R., & Kalyanasundaram, M. (2009). Larvicidal activity of *Saraca indica*, *Nyctanthes arborescens*, and *Clitoria ternatea* extracts against three mosquito vector species. *Parasitology research*, 104, 1017-1025.
 105. Mattingly, P. F., & Knight, K. L. (1956). The mosquitoes of Arabia. *International Bulletin of British Museum and Natural History (Entomology)*, 4(3), 89-141.
 106. Maurya, P., Mohan, L., Sharma, P., Batabyal, L., & Srivastava, C. N. (2007). Larvicidal efficacy of *Aloe barbadensis* and *Cannabis sativa* against the malaria vector *Anopheles stephensi* (Diptera: Culicidae). *Entomological research*, 37(3), 153-156.
 107. Meade, M. S., Florin, J. W., & Gesler, W. M. (1998). *Medical geography*. New York, NY: Guilford Press.
 108. Mehmood, A., Naeem, M., Raza, A. B. M., Majeed, M. Z., Ullah, M. I., Riaz, M. A., ... & Raza, W. (2024). Faunal and habitat distribution of mosquitoes (Diptera: Culicidae) in Chakwal, Punjab, Pakistan. *Sarhad Journal of Agriculture*, 40(2), 463-469.
 109. Miranda, C., Marques, C. C. A., & Massa, J. L. (1998). Satellite remote sensing as a tool for the analysis of the occurrence of American cutaneous leishmaniasis in Brazil. *Revista de Saúde Pública*, 32(5), 455-463.
 110. Moncayo, A. C., Edman, J. D., & Finn, J. T. (2000). Application of geographic information technology in determining risk of eastern equine encephalomyelitis virus transmission.
 111. *Journal of the American Mosquito Control Association*, 16(1), 28-35.
 112. Moretti, R., Lim, J. T., Ferreira, A. G. A., Ponti, L., Giovanetti, M., Yi, C. J., ... & Ross, P. A. (2025). Exploiting Wolbachia as a Tool for Mosquito-Borne Disease Control: Pursuing Efficacy, Safety, and Sustainability. *Pathogens*, 14(3), 285.
 113. Mughini-Gras, L., Mulatti, P., Severini, F., Boccolini, D., Romi, R., Bongiorno, G., & Busani, L. (2014). Ecological niche modelling of potential West Nile virus vector mosquito species and their geographical association with equine epizootics in
 114. Italy. *Ecohealth*, 11, 120-132.
 115. Murty, U. S., Rao, M. S., & Arunachalam, N. (2010). The effects of climatic factors on the distribution and abundance of Japanese encephalitis vectors in Kurnool district of Andhra Pradesh. *Indian Journal of Vector Borne Diseases*, 47(1), 26-32.
 116. Naeem, M., Alahmed, A. M., Khair, S. M., & Sallam, M. F. (2016). Spatial distribution modeling of *Stegomyia aegypti* and *Culex tritaeniorhynchus* (Diptera: Culicidae) in AlBahah Province, Kingdom of Saudi Arabia. *Tropical Biomedicine*, 33(2), 295-310.
 117. Nakhapakorn, K., & Tripathi, N. K. (2005). An information value based analysis of physical and

- climatic factors affecting dengue fever and dengue haemorrhagic fever incidence. *International journal of health geographics*, 4, 1-13.
118. Nansen, C., Campbell, J. F., Phillips, T. W., & Mullen, M. A. (2003). The impact of spatial structure on the accuracy of contour maps of small data sets. *Journal of Economic Entomology*, 96(6), 1617-1625.
 119. Nayab, G. E., Rahman, R. U., Hanan, F., Khan, I., & Fahim, M. (2025). Metagenomic exploration of the bacteriome reveals natural Wolbachia infections in yellow fever mosquito *Aedes aegypti* and Asian tiger mosquito *Aedes albopictus*. *bioRxiv*, 34, 133180.
 120. Omumbo, J., Ouma, J., Rapuoda, B., et al. (1998). Mapping malaria transmission intensity using geographical information systems (GIS): an example from Kenya. *Annals of Tropical Medicine and Parasitology*, 92(1), 7-21.
 121. Ortega-Huerta, M. A., & Townsend, P. A. (2008). Modelling ecological niches and predicting geographic distributions: a test of six presence-only methods. *Revista Mexicana de Biodiversidad*, 79, 205-216.
 122. Palaniyandi, M., & Palaniyandi, M. (2014). Web mapping GIS: GPS under the GIS umbrella for *Aedes* species dengue and chikungunya vector mosquito surveillance and control Rapid Epidemiological Mapping of Lymphatic Filariasis in Southern India View project Application of Remote Sensing and Geographic. *International Journal of Mosquito Research*, 1(3), 18-25.
 123. Parham, P. E., Pople, D., Christiansen-Jucht, C., Lindsay, S., Hinsley, W., & Michael, E. (2012). Modeling the role of environmental variables on the population dynamics of the malaria vector *Anopheles gambiae* sensu stricto. *Malaria Journal*, 11, 271.
 124. Pearson, R. G., Raxworthy, C. J., Nakamura, M., & Peterson, A. T. (2007). Predicting species distributions from small numbers of occurrence records: a test case using cryptic geckos in Madagascar. *Journal of Biogeography*, 34, 102-117.
 125. Peterson, A. T., Soberon, J., & Sanchez-Cordero, V. (1999). Conservatism of ecological niches in evolutionary time. *Science*, 285(5431), 1265-1267.
 126. Pherez, F. M. (2007). Factors affecting the emergence and prevalence of vector borne infections (VBI) and the role of vertical transmission (VT). *Journal of Vector Borne Diseases*, 44(3), 157-163.
 127. Phillips, S. J., & Dudik, M. (2008). Modeling of species distributions with Maxent: new extensions and a comprehensive evaluation. *Ecography*, 31(2), 161-175.
 128. Phillips, S. J., Anderson, R. P., & Schapire, R. E. (2006). Maximum entropy modelling of species geographic distributions. *Ecological modelling*, 190(3-4), 231-259.
 129. Phillips, S. J., Dudik, M., & Schapire, R. E. (2004). A Maximum Entropy Approach to Species Distribution Modelling. *Proceedings of the 21st International Conference on Machine Learning*, 655-662.
 130. Porphyre, T., Bicot, D. J., & Sabatier, P. (2005). Modelling the abundance of mosquito vectors versus flooding dynamics. *Ecological Modelling*, 183, 173-181.
 131. Reisen, W., Lothrop, H. (1999). Effects of sampling design on the estimation of adult mosquito abundance. *Journal of the American Mosquito Control Association*, 15(1), 105-114.
 132. Rejmankova, E., Roberts, D. R., Pawley, A., et al. (1995). Predictions of adult *Anopheles albimanus* densities in villages based on distances to remotely sensed larval habitats. *American Journal of Tropical Medicine and Hygiene*, 53(5), 482-488.
 133. Ritchie, S. A., Hanna, J. N., Hills, S. L., Piispanen, J. P., John, W., McBride, H., Pyke, A., & Spark, R. L. (2002). Dengue control in North Queensland, Australia: case recognition and selective indoor residual spraying. *Dengue Bulletin*, 26, 7-13.
 134. Rochlin, I., Ninivaggi, D. V., Hutchinson, M. L., & Farajollahi, A. (2013). Climate change and range expansion of the Asian tiger mosquito (*Aedes albopictus*) in Northeastern USA: implications for public health practitioners. *PloS One*, 8, 145-180.
 135. Rogers, D. J. (2000). Satellites, space, time and the African trypanosomiasis. *Advances in Parasitology*, 47, 129-171.
 136. Rogers, D. J., Randolph, S. E., Snow, R. W., & Hay, S. I. (2002). Satellite imagery in the study and forecast of malaria. *Nature*, 415, 710-715.
 137. Rohani, A., Wan Najdah, W. M., Zamree, I., Azahari, A. H., Mohd Noor, I., et al. (2010). Habitat characterization and mapping of *Anopheles maculatus* (Theobald) mosquito larvae in malaria endemic areas in Kuala Lipis, Pahang, Malaysia. *Southeast Asian Journal of Tropical Medicine and Public Health*, 41(4), 821-830.
 138. Ryan, P. A., Lyons, S. A., Asemgeest, D., Thomas, P., & Kay, B. H. (2004). Spatial statistical analysis of adult mosquito (*Diptera: Culicidae*) counts: an example using light trap data, in Redland Shire, southeastern Queensland, Australia. *Journal of medical entomology*, 41(6), 1143-1156.
 139. Rydzanicz, K., Hoffman, K., Jawieñ, P., Kiewra, D., & Becker, N. (2011). Implementation of Geographic Information System (GIS) in an environment friendly mosquito control programme in irrigation fields in Wrocław (Poland). *European Mosquito Bulletin*, May 2014, 1-12.
 140. Rydzanicz, K., Lonc, E., Kiewra, D., DeChant, P., Krause, S., & Becker, N. (2009). Evaluation of three microbial formulations against *Culex pipiens pipiens* larvae in irrigation fields in Wrocław, Poland. *Journal of the American Mosquito Control Association*, 25, 140148.
 142. Saleem, A., & Mahmood, S. (2023). Spatio-temporal assessment of urban growth using multistage satellite imageries in Faisalabad, Pakistan. *Journal of Urban and Regional Analysis*, 3(1), 10-18.
 143. Sallam, M. F., Al Ahmad, A. M., Abdel-Dayem, M. S., & Abdullah, M. A. R. (2013). Ecological niche modeling and land cover risk areas for Rift Valley Fever vector *Culex tritaeniorhynchus* Giles in Jazan, Saudi Arabia. *PLoS One*, 8(6).
 144. Siria, D. J., Batista, E. P. A., Opiyo, M. A., Melo, E. F., Sumaye, R. D., Ngowo, H. S., Eiras, A. E., & Okumu, F. O. (2018). Evaluation of a simple polytetrafluoroethylene (PTFE) based membrane for blood-feeding of malaria and dengue fever vectors in the laboratory. *Parasites & Vectors*, 11(1), 1-10.
 145. Smith, L. B., Kasai, S., & Scott, J. G. (2016). Pyrethroid resistance in *Aedes aegypti* and *Aedes albopictus*: Important mosquito vectors of human diseases. *Pesticide Biochemistry and Physiology*, 133, 1-12.
 146. Smith, T., Charlwood, J. D., Takken, W., Tanner, M., & Spiegelhalter, D. J. (1995). Mapping the densities of malaria vectors within a single village. *Acta Tropica*, 59(1), 1-18.
 147. Su, M. D. (1994). Framework for application of geographic information system to the monitoring of dengue vectors. *Kaohsiung Journal of Medical Science*, 10, 94-101.
 148. Surveillance report. (2007). Jeddah, Saudi Arabia, Ministry of Health, Department of Communicable

Diseases.

149. Tedrow, C. A. (2010). *Using remote sensing, ecological niche modeling, and geographic information systems for Rift Valley fever risk assessment in the United States* (Doctoral dissertation, George Mason University).
150. Thomson, M. C., Connor, C. J., Milligan, P., et al. (1997). Mapping malaria risk in Africa: what can satellite data contribute? *Parasitology Today*, 13(8), 313–318.
151. Thomson, M. C., Connor, S. J., Milligan, P. J. M., & Flasse, S. P. (1996). The ecology of malaria—as seen from Earth-observation satellites. *Annals of Tropical Medicine & Parasitology*, 90(3), 243–264.
152. Thomson, M. C., Elnaïem, D. A., Ashford, R. W., & Connor, S. J. (1999). Towards a kala azar risk map for Sudan: mapping the potential distribution of *Phlebotomus orientalis* using digital data of environmental variables (vol 4, pg 105, 1999). *Tropical Medicine & International Health*, 4(3), 240–241.
153. Tucker, C. J., & Nicholson, S. E. (1999). Variations in the size of the Sahara Desert from 1980–1997. *AMBIO: A Journal of the Human Environment*, 28, 587–591.
154. Ullah, U. N., Hafeez, F., Ali, S., Arshad, M., Akram, W., Ali, A., ... & AIMunqedhi, B. M. (2023). Distribution of mosquito species in various agro-ecological zones of Punjab. *Journal of King Saud University-Science*, 35(8), 102874.
155. Vanek, M. J., Shoo, B., Mtasiwa, D., Kiama, M., Lindsay, S. W., Fillinger, U., Kannady, K., Tanner, M., & Killeen, G. F. (2006). Community-based surveillance of malaria vector larval habitats: a baseline study in urban Dar es Salaam, Tanzania. *BMC Public Health*, 6, 154.
156. Veerakumar, K., & Govindarajan, M. (2014). Adulticidal properties of synthesized silver nanoparticles using leaf extracts of *Feronia elephantum* (Rutaceae) against filariasis, malaria, and dengue vector mosquitoes. *Parasitology Research*, 113(11), 4085–4096.
157. WHO, L. F. (1992). *The Disease and Its Control: 5th Report of the WHO Expert Committee on Filariasis* (No. 821, p. 56). Technical Report Series.
158. Wiley, E. O., McNyset, K. M., Peterson, A. T., Robins, C. R., & Stewart, A. M. (2003). Niche modeling and geographic range predictions in the marine environment using a machinelearning algorithm. *Oceanography*, 16, 120–127.
159. Wilke, A. B. B., & Marrelli, M. T. (2015). Paratransgenesis: A promising new strategy for mosquito vector control. *Parasites and Vectors*, 8(1), 1–9.
160. Wisz, M. S., Hijmans, R. J., Li, J., Peterson, A. T., Graham, C. H., Guisan, A., Group NPSDW. (2008). Effects of sample size on the performance of species distribution models. *Diversity and Distributions*, 14, 763–773.
161. Wood, B. L., Beck, L., Washino, R., et al. (1992). Estimating high mosquito-producing rice fields using spectral and spatial data. *International Journal of Remote Sensing*, 13(15), 2813–2826.
162. Wood, B. L., Washino, R., Palchick, S., et al. (1991b). Spectral and spatial characterization of rice field mosquito habitat. *International Journal of Remote Sensing*, 12(4), 621–626.
163. Wood, B., Washino, R., Beck, L., et al. (1991a). Distinguishing high and low *anopheline* producing rice fields using remote sensing and GIS technologies. *Preventive Veterinary Medicine*, 11(4), 277–288.

Disclaimer / Publisher's Note

The statements, opinions, and data contained in all publications of the *PAKISTAN JOURNAL OF ZOOLOGICAL SCIENCES (PJZS)* are solely those of the individual author(s) and contributor(s) and do not necessarily reflect those of IJSMART Publishing and/or the editor(s). IJSMART Publishing and/or the editor(s) disclaim any responsibility for any injury to persons or property resulting from any ideas, methods, instructions, or products mentioned in the content.